

LAMP-BART

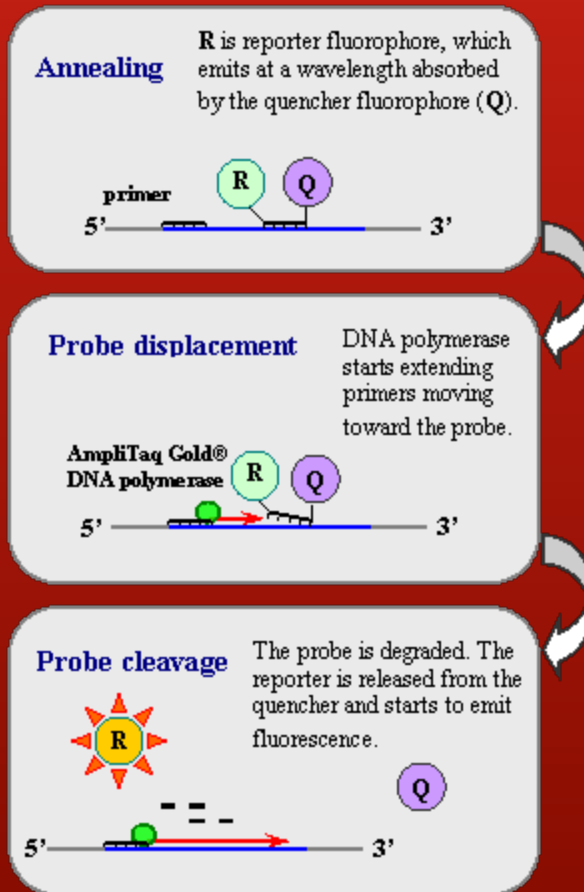
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Current molecular diagnostic system

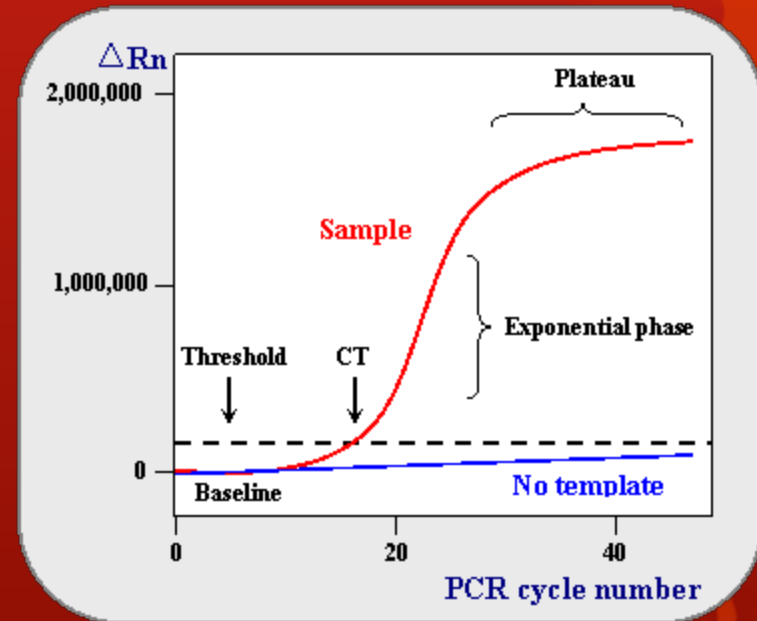
- Detection of target nucleic acid
- quantitative PCR (qPCR)
- Fluorescent dye/fluorescently-labelled oligonucleotides

Quantitative PCR (qPCR)

TaqMan® Applied Biosystems



Model of real time quantitative PCR plot



NCBI 2012. Published online:
<http://www.ncbi.nlm.nih.gov/projects/genome/probe/doc/TechQPCR.shtml>

Quantitative PCR (qPCR)

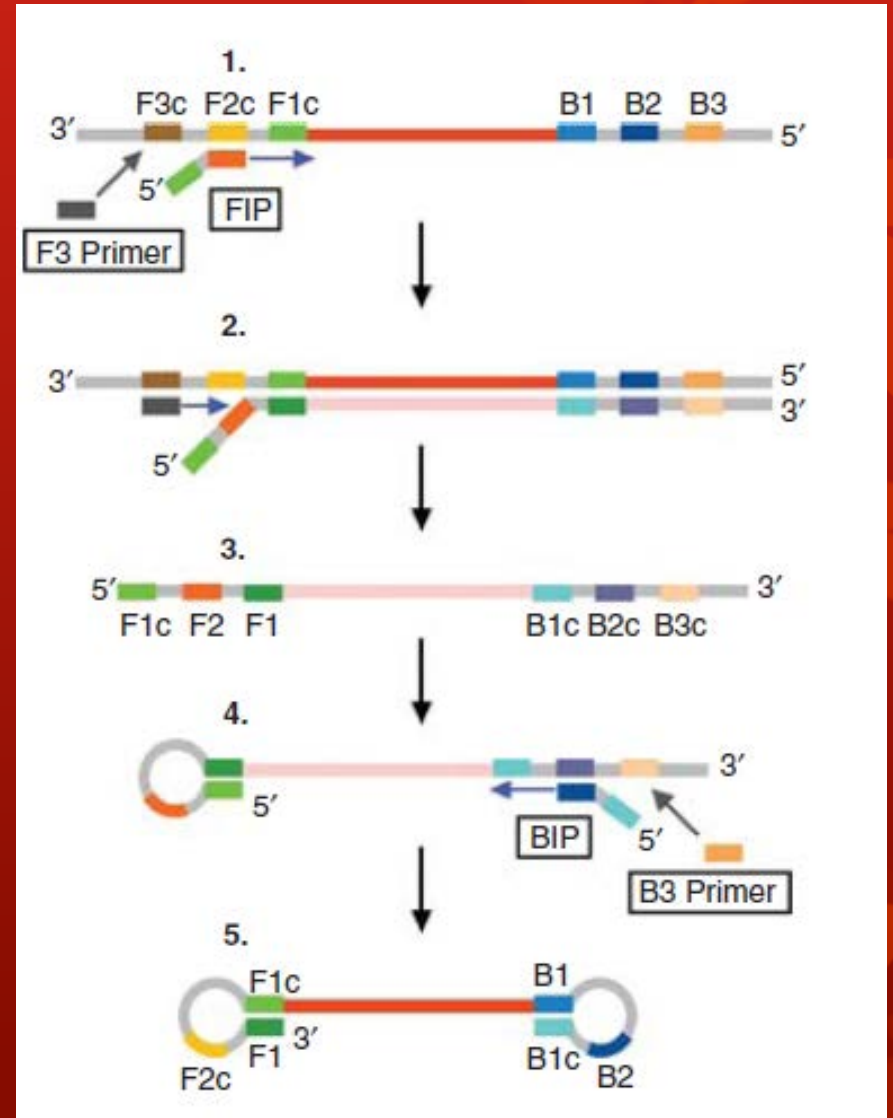
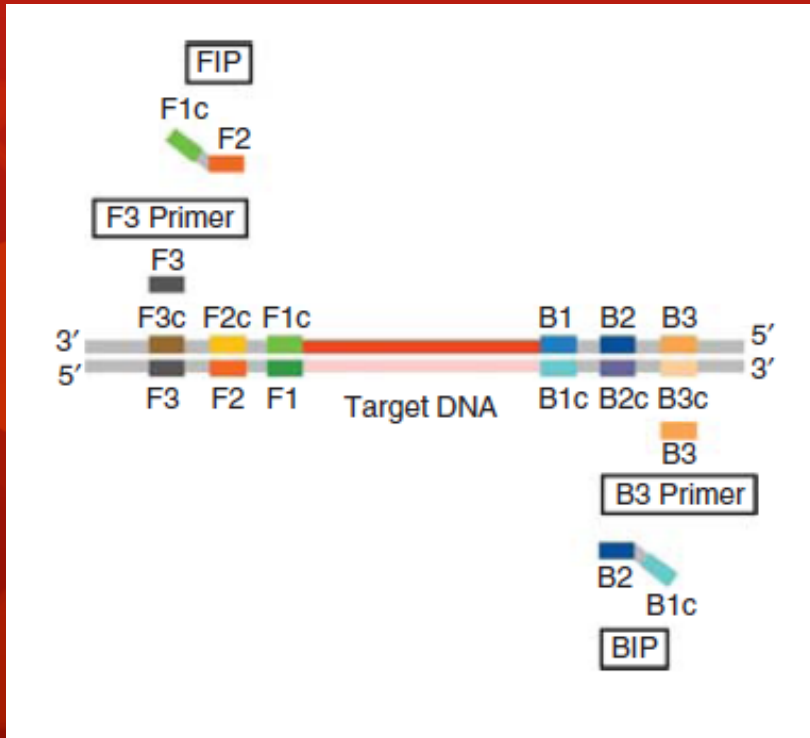
- Strict requirements on assay equipments
 - Temperature cycling
 - Wavelength-specific fluorescent excitation
 - Emission measurement
- Limitations
 - Power consumption
 - Optical arrangements required
- Production of low-cost, simple and robust instruments

LAMP-BART

- LAMP
- Loop-mediated amplification
- Alternative amplification method
- Strand-displacing polymerase
- Isothermal amplification ($\leq 65^{\circ}\text{C}$)
- 4 primers recognize 6 regions

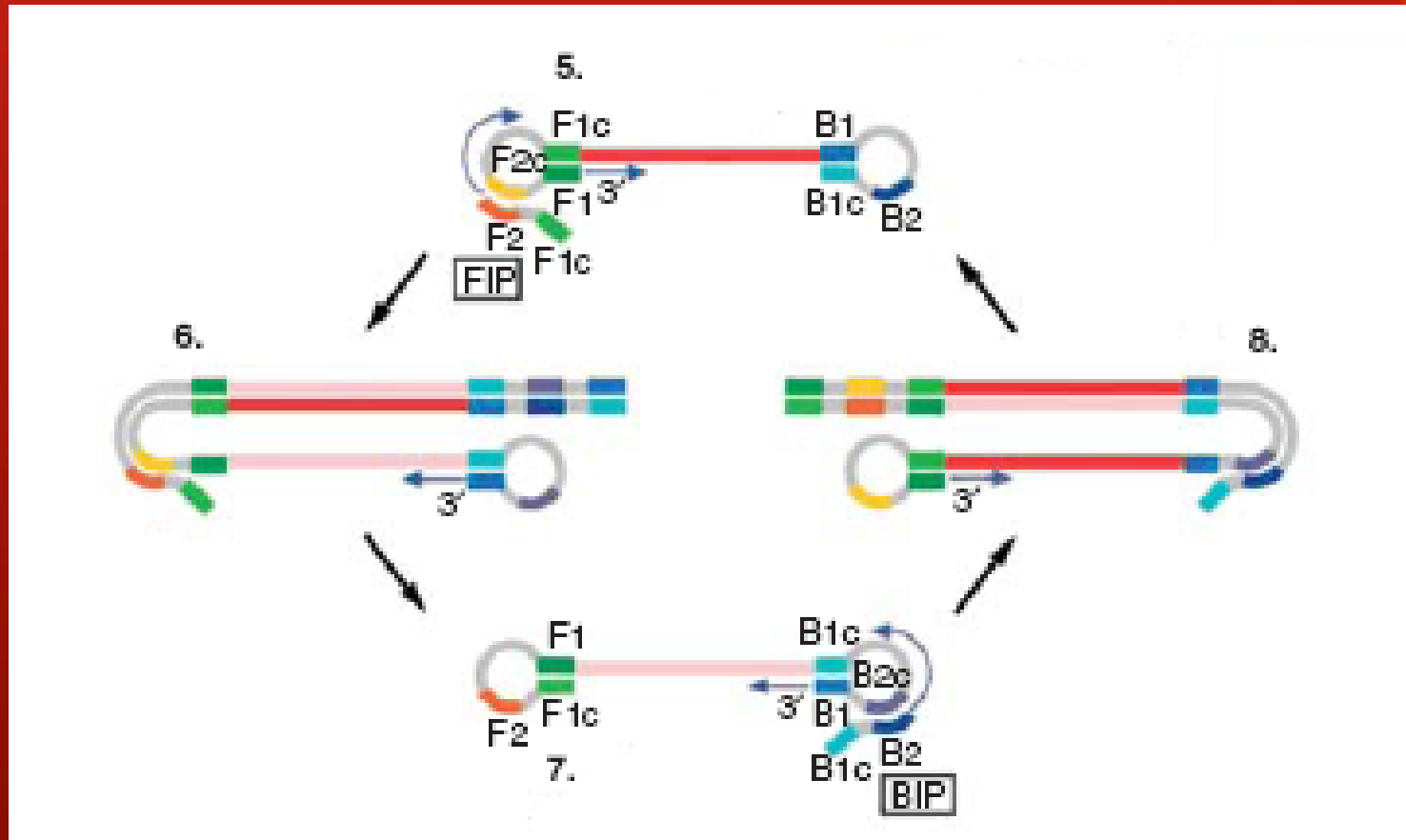
LAMP

1. Starting-structure producing step



LAMP

2. Cycling amplification step



LAMP-BART

- BART
- Bioluminescent Assay in Real-Time
- Alternative amplification detection method
- Inorganic pyrophosphate (PP_i)
- By-product of all amplification

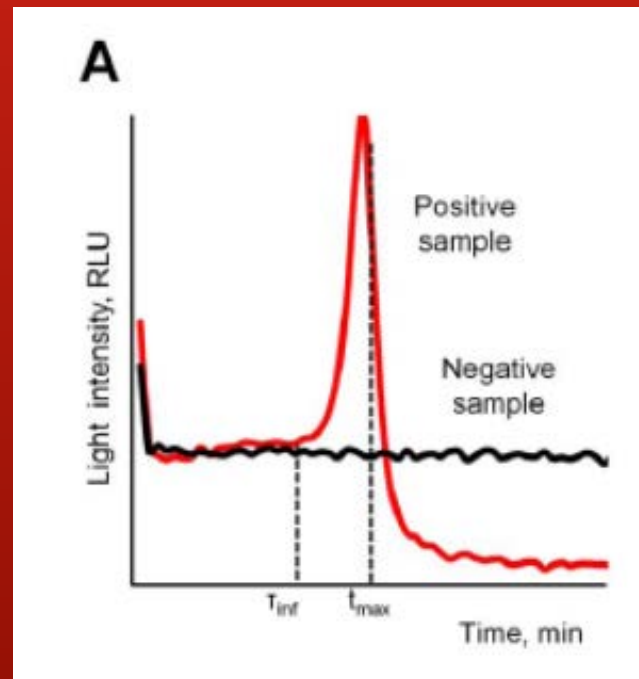
BART



Gandelman *et al* (2010)

- 1 molecule released each nucleotide addition
- Proportional to amount of polynucleotide synthesized
- Starting template concentration

LAMP-BART

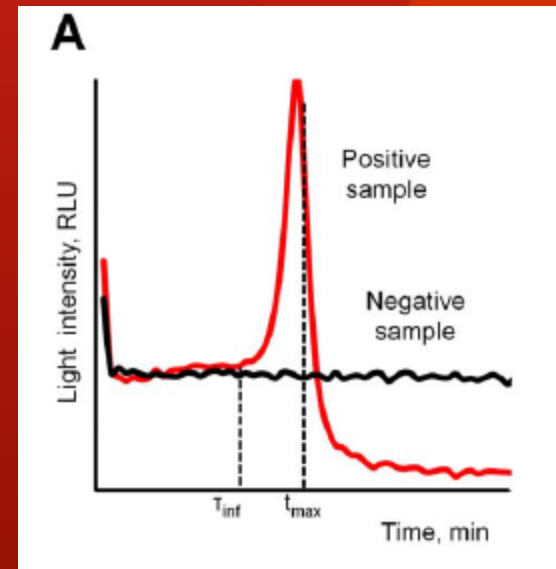


Gandelman *et al* (2010)

55°C 1hr

LAMP-BART

- qPCR curve vs BART curve
- Sigmoidal curve vs sharp peak
- Rapid reduction in bioluminescence after the peak
 - APS exhausted
 - PPi inhibits luciferase



DNA polymerase (1)

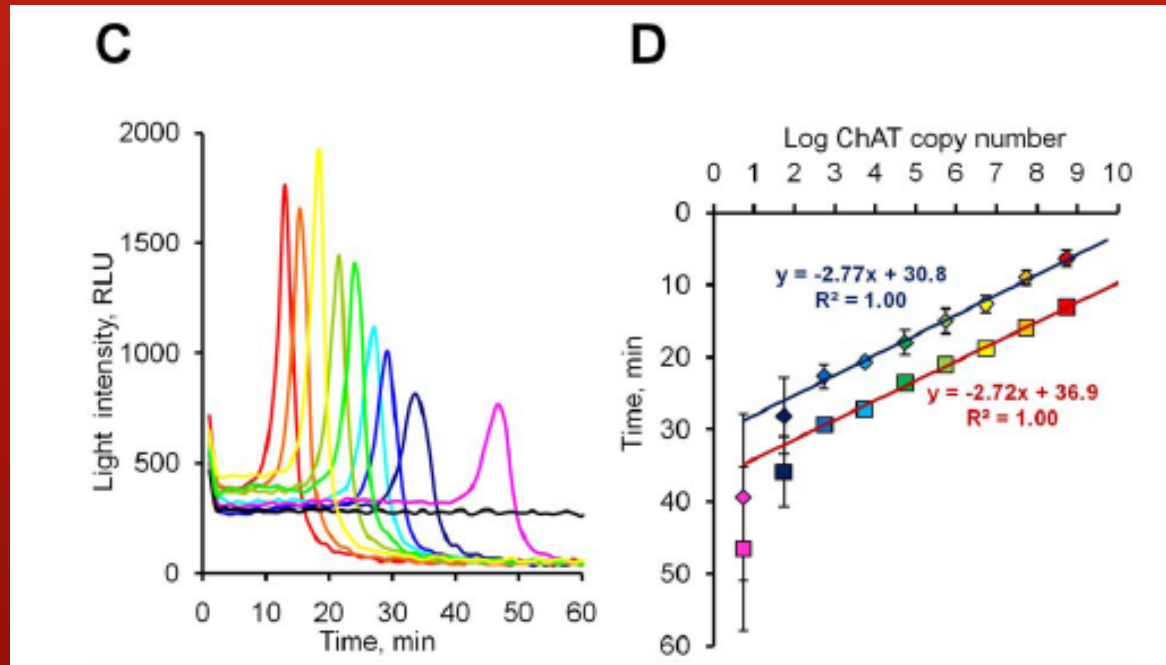


ATP sulfurylase (2)



Firefly luciferase (3)

Quantitative LAMP-BART



Gandelman *et al* (2010)

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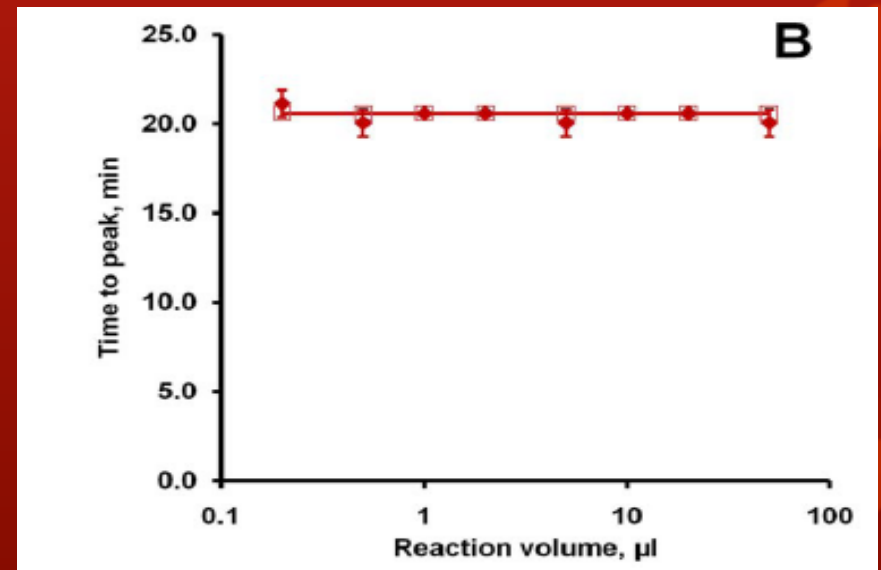
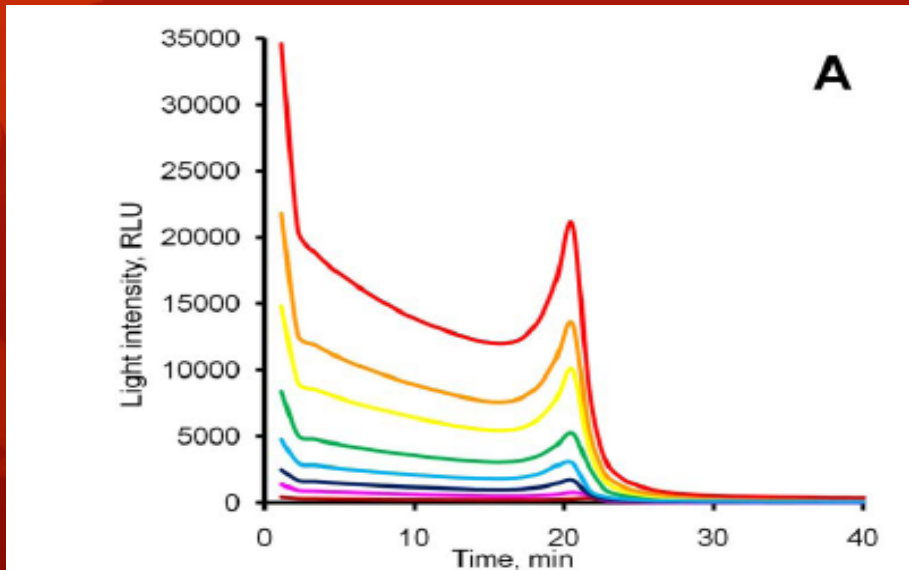
Linear correlation, similar to correlation of Ct value to DNA template load in qPCR

Quantitative LAMP-BART

- Time required for same amount PP_i to be released = amount of target nucleic acid
- Time-to-peak α starting target gene conc.

Quantitative LAMP-BART

- Amplification with same DNA conc. but different reaction volume
- Light intensity, reduction in volume
- Time-to-peak unchanged
- Depend on time-to-peak, not absolute light intensity output



Application of LAMP-BART

- Detection of *Chlamydia trachomatis* (CT) in clinical specimens
- 105 clinical urine specimens of unknown CT status
- Bacterial DNA isolated
- Compared with qPCR result

Application of LAMP-BART

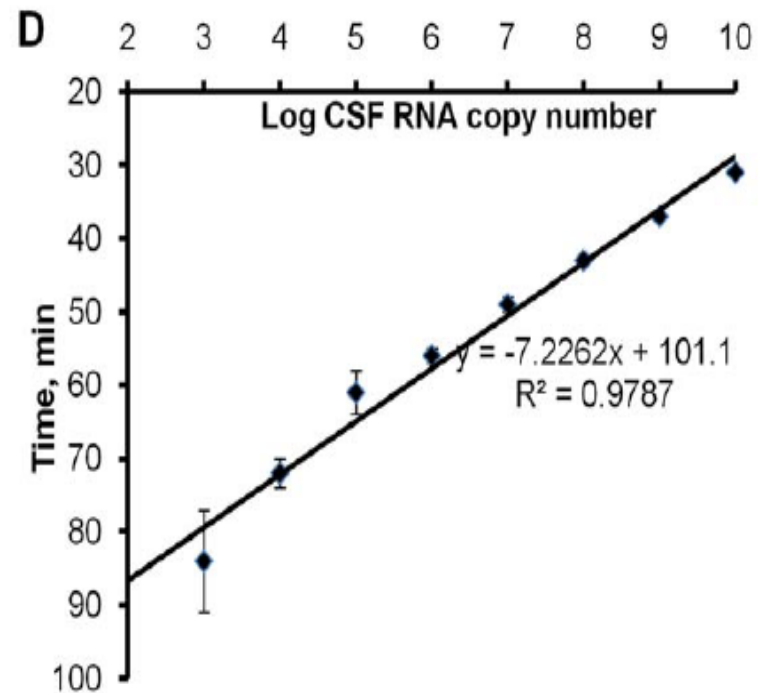
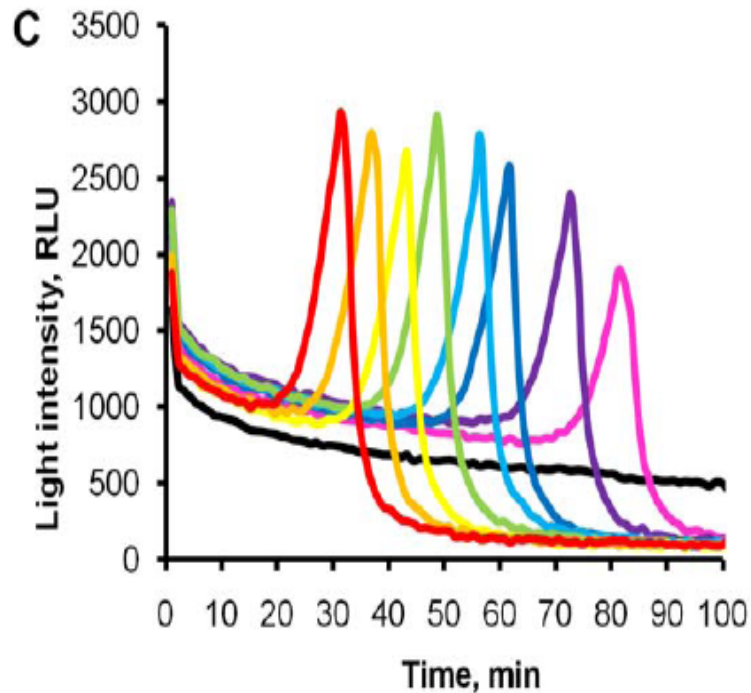
	LAMP-BART	qPCR
Total number of samples	105	105
CT-positive samples	43 ($t_{\max} < 60$ min)	45 ($Ct \leq 40$ cycles)
CT-negative samples	62	60 ($Ct > 40$)
Sensitivity, %	95.6	100
Specificity, %	100	100
Assay time	60 min	2.5 hours
Mean t_{\max} or Ct/equivalent time	33.6 min	35.2 cycles \sim 1 h 46 min

Gandelman *et al* (2010)

Application of RT-LAMP-BART

- Classical swine fever virus
- Detection of RNA template
- Purified RNA amplified in closed-tube one-step format
- Reverse transcription, LAMP amplification, BART detection reagents

Application of RT-LAMP-BART

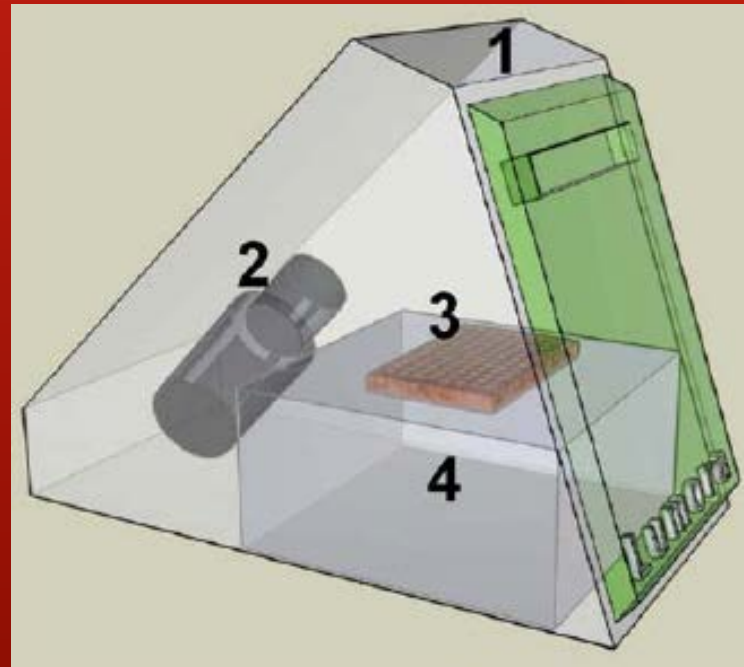


Gandelman *et al* (2010)

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Coupled RT-LAMP-BART quantification and detection of viral RNA genome for diagnostics

Instruments for LAMP-BART



Advantages of LAMP-BART

- Coupled amplification and detection system in single closed tube
- Reduce risk of contamination
- Effectively used on patient-derived samples, tolerance to inhibitors in patient sample
- Straightforward, simple, rapid
- LAMP: 4 primers, 6 recognition sites, high specificity
- Cost-effective
 - Simple light detector
 - No need temperature cycle, wavelength-specific fluorescent excitation, emission measurement
 - Require only a constant temp. maintained by heat block
- Quantitation rely on time-to-peak but not absolute intensity
 - Tolerance to contaminating ATP or PP_i
 - Rate of change not absolute level determined

Possible drawback of LAMP-BART

- Single signal, not possible for multiplex PCR

Conclusion

- Novel coupled amplification and detection
- Single close-tubed format
- Simple, fast, cost-effect quantitative assay

References

- NCBI. 2012. Published online:
<http://www.ncbi.nlm.nih.gov/projects/genome/probe/doc/TechQPCR.shtml>
- Tomita *et al.* 2008. Loop-mediated isothermal amplification (LAMP) of gene sequences and simple visual detection of products. *Nature Protocols* 3, - 877 - 882
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- Kiddle *et al.* 2012. GMO detection using a bioluminescent real time reporter (BART) of loop mediated isothermal amplification (LAMP) suitable for field use. *BMC Biotechnol.* 30;12:15